

# The role of astrocyte alterations in early changes in the dynamics of cultured cerebellar networks

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An aberrant response to DNA lesions is implicated in many human brain degenerative disorders. Various types of DNA lesions activate a cellular process known as the DNA damage response (DDR). Mutations affecting the proteins involved in the DDR can lead to severe genomic instability syndromes that involve varying degrees of sensitivity to genotoxic stress, and also to tissue degeneration, cancer predisposition, and premature aging. Malfunctioning DDR was found in various brain degenerative disorders such as Alzheimer's, Parkinson's and Huntington. One of the key components of the DDR is the protein ATM, which is inactivated in the genomic instability disorder ataxia-telangiectasia (A-T). In order to study the effect of malfunctioning DDR on neuronal circuits, we used calcium imaging and immunocytochemical staining to compare the morphology and the dynamics of primary cerebellar cultures grown from postnatal *Atm*-deficient and wild-type (WT) mice. Cerebellar networks exhibited spontaneous network events after two weeks *in-vitro*. Compared to WT circuits, *Atm*-deficient circuits displayed a lower number of global synchronizations and a larger number of sparse synchronizations, i.e. synchronous events involving less than a dozen cells. In WT networks we observed significantly high global burst similarity compared to the *Atm*<sup>-/-</sup> network. In addition, nodes with a high functional connectivity degree could be observed in the WT networks but not in the *Atm*<sup>-/-</sup> networks. To understand A-T on the cellular level we tested the hypothesis that A-T is at least partially a glial disease. Immunocytochemical staining of astrocytes revealed a significantly less complex cell arborization in *Atm*-deficient versus WT circuits, as measured by the number of branches originating from cell bodies as well as their length. To further study the interrelations between neurons and astrocytes, we generated chimeric networks in which the neurons and astrocytes were extracted from different animals. We found that functional and viable chimera cultures could be prepared only from P8 cerebellar neurons and astrocytes. Chimera cultures made from combinations of P8 cerebellar neurons and P2 cortical glia or from P8 cerebellar neurons and P2 cerebellar glia did not survive and the neurons died within 3 to 4 days of plating. Our results clearly show that *Atm*<sup>-/-</sup> astroglial cell replacement with WT astrocytes fully restores the dynamics of neural networks in chimera neuron-glia networks extracted from *Atm*-deficient mice. In contrast, *Atm*<sup>-/-</sup> astrocytes failed to support the survival and the functionality of the WT neurons. These results support the notion that neuronal network failures in genetic brain degenerative diseases are correlated with impairment of astroglial cell functionality.